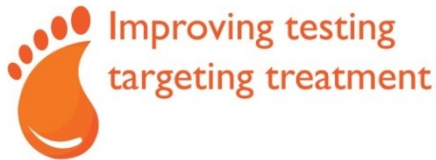


Applications of Next Generation Sequencing for Newborn Screening

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The University Of Sheffield.

Expanded Newborn Screening

CLAHRC for South Yorkshire



National Institute for Health Research



Department of Health

Health Innovation Challenge Fund

Overview of Presentation

- Challenges and advantages of Next Generation Sequencing (NGS) for NBS
- Project aims, progress made and future work
 - 1) Genotype-phenotype database development
 - 2) Laboratory optimisation of NGS for NBS

Challenges of NGS for NBS

- Hot topic – concerns regarding whole genome or whole exome sequencing for NBS
- Medical, psychological, ethical and economic reasons against
- Need clear best practise implementation guidelines
- Should focus on targeted gene analysis only?

The potential role of NGS for NBS

- Primary test for current disorders
- Primary test for disorders where there is no suitable biochemical test available
- Adjunct test to biochemical testing – use in combination to confirm metabolic results
- Obtain a clearer picture of disease severity, expected onset (early or late) and treatment regimen

Aim 1: database development

- Development of a genotype-phenotype database
- Important clinical resource
- Enhance disease understanding

- For 6 inherited metabolic diseases:
MSUD, HCU, IVA, GA1, PKU & MCADD

Current Data Fields

Participant Information
Birth, Development and Growth
Genetic Sequencing Results

Screen Identified
Initial diagnostic visit

Screen Identified
Regular visit

Clinically Identified
Initial diagnostic visit

Clinically Identified
Regular visit

NBS data

Medication, Diet, other advice;
Biochemical tests; Symptoms

Medication, Diet, other advice;
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Medication, Diet, other advice;
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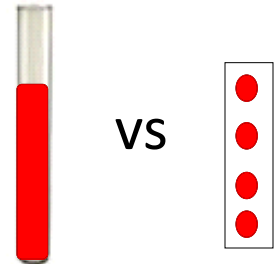
Medication, Diet, other advice;
Biochemical tests; Symptoms

NGS and Reporting

- Sequencing 150 healthy control and 260 patient samples
- 150 healthy controls and 125 patients recruited so far
- Ampliseq Panel Design for six NBS disorders (13 genes)
- Validating specificity and sensitivity of variant calling
- Diagnostic reporting for all patients recruited
- Genetic variant information transferred to project database

Aim 2: Optimisation of High-Throughput NGS for NBS

- Utilise healthy control individuals' DNA
- Compare DNA extracted from Venous Blood (VB) with DNA extracted from Dried Blood Spots (DBS)
- Aim to obtain same sequence quality from DBS DNA as from VB DNA using NGS
- Use current screened disorders to trial the analysis



Laboratory Work Overview

DNA extraction: DBS and VB samples



Assess DNA quality and quantity for NGS



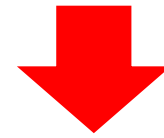
Design NBS disorder gene panel



Library to sample preparation for NGS

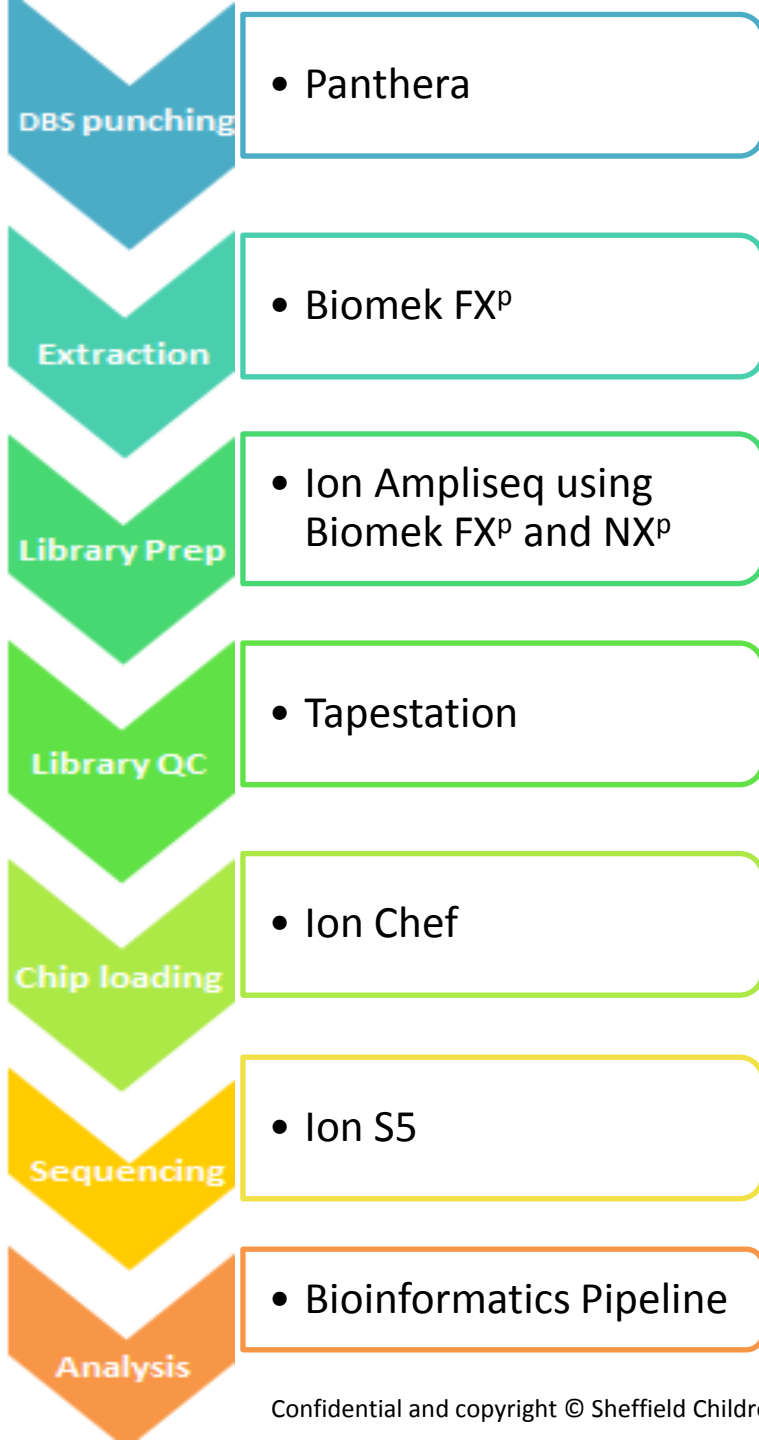


Validation of method for NBS: automation, speed and cost



DNA Extraction Results

- A small sample size of healthy control DNA used to optimise extraction methods:
 - 2 variations of VB extraction
 - 10 variations of DBS extraction
- Sufficient quality and quantity of DNA extracted from DBS for use in NGS
 - No significant loss of variant information between VB and DBS DNA
 - Method chosen for DBS extraction that is cost effective and can be automated



Automated Workflow

Booking on and Punching

- Booking on of guthrie cards at Sheffield NBS Facility
- Use Panthera DBS puncher with 6 mm punch head
- Punching time: 60 minutes per 96-well barcoded plate
- SDGS will book on 96-well plates containing DBS samples

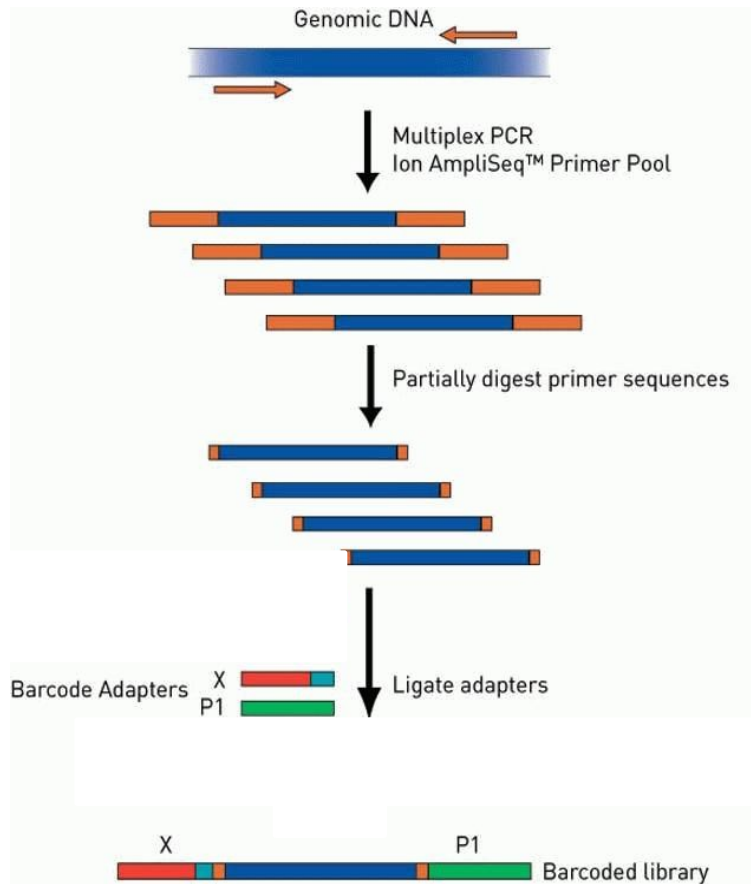


DNA Extraction

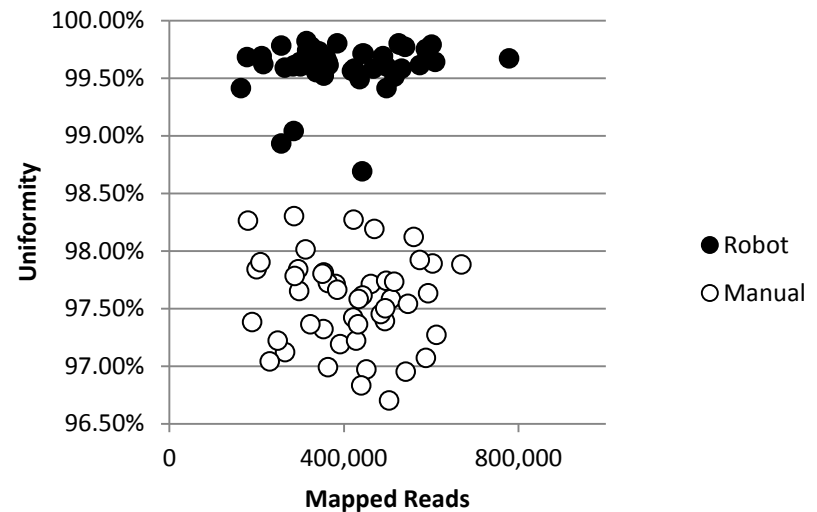
- Biomek FX^P robot program developed
- 3x 96-well plates in 90 minutes
- Sequencing quality of DBS is similar to VB samples



Ampliseq Library Preparation



- Semi-Automated method on the Biomek FX^p robot
- Robot outperforms manual library preparation in terms of uniformity of coverage ($p < 0.04$)



Chip Loading and Sequencing

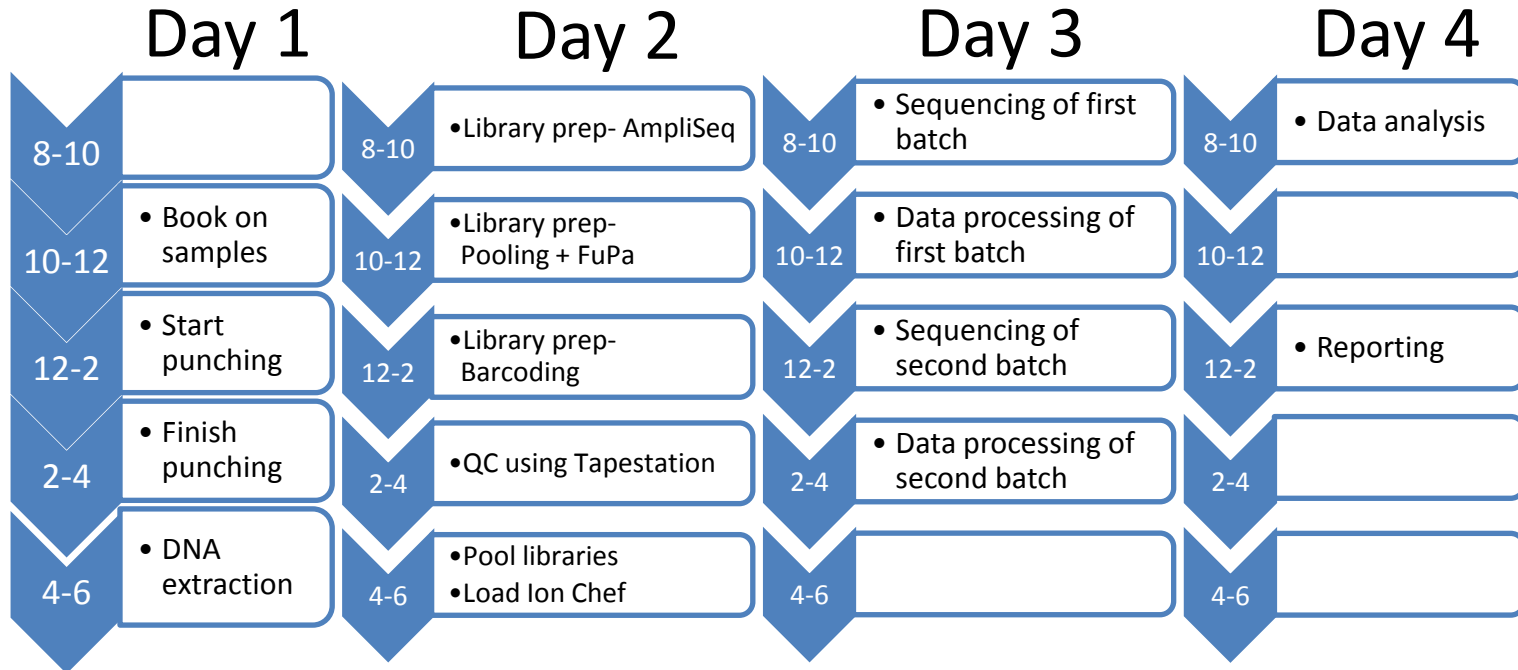


- Ion Chef: 15 min hands on time; 11h run time (overnight); 2x chips per run
- 192 samples per run using 2x Ion 540 chips
- Ion S5: <15 min hands on time; 2.5h run time per chip
- No significant difference in sequencing coverage or variant calling between DBS and VB samples
- High-throughput bioinformatics analysis pipeline

Other Laboratory Progress

- Validated 2 NBS panels according to ACGS Guidelines:
“Best practice guidelines for Targeted Next Generation Sequencing Analysis and Interpretation, 2015”
- Pathogenic variants can be identified using patient sequencing data
- There is no significant effect of the DBS age (1, 7, 30 and 60 days) on sequencing quality and variant calling
- Discussions with Thermo Fisher and Perkin Elmer are helping to drive laboratory developments

Current Timeline for NBS

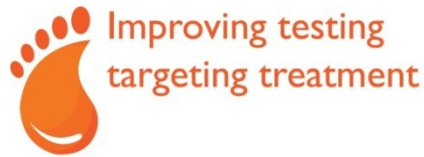


- 2 x 96 samples would be punched per day meaning that ~ 1000 samples could be sequenced per week.
- Doubling up on automation equipment would increase sample high-throughput capability.

On-going work

- Aim1: Database Development
 - Continue sequencing healthy control and patient VB samples
 - Collect patient medical record information into database
 - Perform genotype-phenotype correlative analysis
- Aim 2: Development of high-throughput NGS for NBS:
 - Validate specificity and sensitivity of NGS process
 - Test high-throughput capability and turnaround times of the whole process from booking on to NGS reporting
 - Confirm proof of concept: 1000-2000 samples in one week
 - Perform cost-benefit analysis

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Department of Health

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