



OVERVIEW OF METHODS FOR DETERMINATION OF ACYLCARNITINES IN SERUM OR PLASMA

This document summarizes a number of diagnostic tests commonly used to analyse acylcarnitines (AC) in serum or plasma to the diagnosis of inborn errors of metabolism.

Detailed protocols are not included here; these can be found in the literature references provided.

Acylcarnitine analysis has proven a useful tool in the evaluation of patients at risk for inborn errors of fatty acid oxidation and for organic acidemias that are primarily due to defects in branched-chain amino acid metabolism (Matern 2014). Given the diverse clinical presentation of these conditions, AC analysis has become an integral part of the biochemical genetic laboratory investigation of a large number of patients (Carpenter and Wiley 2002), (Millington, Norwood et al. 1989), (Vreken, van Lint et al. 1999).

Analysis in serum or plasma allows for using the same sample taken for other determinations that contribute to the diagnosis or monitoring of metabolic diseases (Van Hove, Kahler et al. 2000), (Delolme, Vianey-Saban et al. 1997).

AC analysis is almost exclusively performed by tandem mass spectrometry (MS/MS) using stable isotope labeled internal standards that allow quantitation of AC species (Matern 2008). This technique has been displacing others in the last years, like GC/MS (Lowes, Rose et al. 1992), (Jones and Chalmers 1995), (Costa, Struys et al. 1997).

Basically, all the methods include an extraction from the sample by mixing with methanol or acetonitrile to precipitate and separate the proteins (Liu and Pasquali 2005), and the addition of isotopically labeled AC to be used as internal standards. Following steps vary depending on the possibility of derivatization, chromatographic separation and methods to quantify (Ruiz-Sala, Ruijter et al. 2016).

The improvement of sensitivity of the newest mass spectrometers has allowed the analysis of the free AC without derivatization (Ghoshal, Guo et al. 2005).

Derivatization as butyl esters was used to gain sensitivity in the first mass spectrometers, to ensure the positive charge of the AC. Also the use of methyl esters has been reported; however, the identification of the analytes could be more difficult.

Flow injection analysis (FIA) is the more commonly use procedure to start the analysis, since the run times are shortened and also simplify the quantification of the AC. However, this kind of injection does not consider the separation of isomers, that sometimes is essential for diagnosis, avoiding more diagnostic tests (Abdenur, Chamoles et al. 1998), (Silva, Selhorst et al. 2001). The insertion of a liquid chromatographic column solves this problem (Ferrer, Ruiz-Sala et al. 2007), (Gaskell, Guenat et al. 1986).



Tandem mass spectrometry acquisition modes are mainly two: Precursor ion scanning of m/z 85, which detects all compounds with this fragment, or MRM (multiple reaction monitoring) to improve sensitivity of a selected number of AC. Also precursor ion of m/z 99 is used (Millington and Stevens).

Finally, different quantification methods can be employed. Software used in newborn screening is also applicable in FIA, it is fast and simple (Rashed, Ozand et al. 1995). The introduction of calibration curves improves quantitative parameters as accuracy or linearity of response (Minkler, Stoll et al.).

Document prepared by:

Pedro Ruiz-Sala
PhD in Pharmacy
Centro de Diagnóstico de Enfermedades Moleculares
Universidad Autónoma de Madrid
28049 Madrid
Email: prsala@cbm.csic.es

-
- Abdenur, J. E., N. A. Chamoles, et al. (1998). "Diagnosis of isovaleric acidaemia by tandem mass spectrometry: false positive result due to pivaloylcarnitine in a newborn screening programme." *J Inherit Metab Dis* 21(6): 624-30.
- Carpenter, K. H. and V. Wiley (2002). "Application of tandem mass spectrometry to biochemical genetics and newborn screening." *Clin Chim Acta* 322(1-2): 1-10.
- Costa, C. G., E. A. Struys, et al. (1997). "Quantitative analysis of plasma acylcarnitines using gas chromatography chemical ionization mass fragmentography." *J Lipid Res* 38(1): 173-82.
- Delolme, F., C. Vianey-Saban, et al. (1997). "[Study of plasma acylcarnitines using tandem mass spectrometry. Application to the diagnosis of metabolism hereditary diseases]." *Arch Pediatr* 4(9): 819-26.
- Ferrer, I., P. Ruiz-Sala, et al. (2007). "Separation and identification of plasma short-chain acylcarnitine isomers by HPLC/MS/MS for the differential diagnosis of fatty acid oxidation defects and organic acidemias." *J Chromatogr B* 860(1): 121-6.
- Gaskell, S. J., C. Guenat, et al. (1986). "Differentiation of isomeric acylcarnitines using tandem mass spectrometry." *Anal Chem* 58(13): 2801-5.



- Ghoshal, A. K., T. Guo, et al. (2005). "Rapid measurement of plasma acylcarnitines by liquid chromatography-tandem mass spectrometry without derivatization." *Clin Chim Acta* 358(1-2): 104-12
- Jones, M. G. and R. A. Chalmers (1995). "Analysis of acylcarnitines by gas chromatography-electron impact mass spectrometry." *Biochem Soc Trans* 23(4): 634S.
- Liu, A. and M. Pasquali (2005). "Acidified acetonitrile and methanol extractions for quantitative analysis of acylcarnitines in plasma by stable isotope dilution tandem mass spectrometry." *J Chromatogr B Analyt Technol Biomed Life Sci* 827(2): 193-8.
- Lowes, S., M. E. Rose, et al. (1992). "Identification of urinary acylcarnitines using gas chromatography-mass spectrometry: preliminary clinical applications." *J Chromatogr* 577(2): 205-14.
- Matern, D. (2008). *Acylcarnitines, Including In Vitro Loading Tests. Laboratory guide to the methods in biochemical genetics.* D. M. Blau N, Gibson LM. Berlin, Heidelberg, Springer Verlag: 171-206.
- Matern, D. (2014). *Acylcarnitines. Physician's guide to the diagnosis, treatment, and follow-up of inherited metabolic diseases.* G. K. Blau N Duran M, Dionisi-Vici C. Berlin Heidelberg, Springer: 775-784.
- Millington, D. S., D. L. Norwood, et al. (1989). "Application of fast atom bombardment with tandem mass spectrometry and liquid chromatography/mass spectrometry to the analysis of acylcarnitines in human urine, blood, and tissue." *Anal Biochem* 180(2): 331-9.
- Millington, D. S. and R. D. Stevens (2011) "Acylcarnitines: analysis in plasma and whole blood using tandem mass spectrometry." *Methods Mol Biol* 708: 55-72.
- Minkler, P. E., M. S. Stoll, et al. (2015) "Validated method for the quantification of free and total carnitine, butyrobetaine, and acylcarnitines in biological samples." *Anal Chem* 87(17): 8994-9001.
- Rashed, M. S., P. T. Ozand, et al. (1995). "Diagnosis of inborn errors of metabolism from blood spots by acylcarnitines and amino acids profiling using automated electrospray tandem mass spectrometry." *Pediatr Res* 38(3): 324-31.
- Ruiz-Sala, P., G. Ruijter, et al. (2016). "Pilot Experience with an External Quality Assurance Scheme for Acylcarnitines in Plasma/Serum." *JIMD Rep* 30: 23-31.
- Silva, M. F., J. Selhorst, et al. (2001). "Characterization of plasma acylcarnitines in patients under valproate monotherapy using ESI-MS/MS." *Clin Biochem* 34(8): 635-8.
- Van Hove, J. L., S. G. Kahler, et al. (2000). "Acylcarnitines in plasma and blood spots of patients with long-chain 3-hydroxyacyl-coenzyme A dehydrogenase deficiency." *J Inherit Metab Dis* 23(6): 571-82.
- Vreken, P., A. E. van Lint, et al. (1999). "Rapid diagnosis of organic acidemias and fatty-acid oxidation defects by quantitative electrospray tandem-MS acyl-carnitine analysis in plasma." *Adv Exp Med Biol* 466: 327-37.